

Adenopure™

ADENOVIRUS PURIFICATION KIT

INSTRUCTION MANUAL



Puresyn, Inc.

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Dear Adenopure™ Kit User,

Thank you for choosing Puresyn, Inc. as your supplier for separation and purification products. Puresyn is dedicated to developing, manufacturing, and marketing state-of-the-art products for the purification of viruses, nucleic acids, and other biological molecules. Since our inception, Puresyn has performed over 400 purification processes for more than 175 companies in the major gene delivery vehicles: supercoiled plasmids, synthetic oligonucleotides, recombinant adenovirus, and adeno-associated virus.

You have chosen an elite product based on Puresyn's proprietary buffer formulations. The buffer formulations contained in this Adenopure™ Kit produce superior results, demonstrated in the Scientific Support section of this manual. This advanced Adenovirus Purification Kit produces ultrapure product. We guarantee you will be satisfied with the Adenopure™ Kit. If you do not agree that this Kit is efficient, convenient, and easy to use, call us at 888-245-9182.

We stand ready to help in all your scientific endeavors. Feel free to contact us with any questions or comments you have.



Dennis M. Flynn
President

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INTRODUCTION AND BACKGROUND

Puresyn's Adenopure™ Kit couples membrane adsorber technology, a simple sample loading mechanism and proprietary buffer formulations to provide rapid and simple isolation of highly purified adenovirus preparations suitable for *in vitro* biological assays.

Membrane adsorber modules are based on a technology that attaches ion exchange functional groups to the inner surface of synthetic microporous membranes in a syringe-filter format.

Membrane adsorber ion exchange modules provide many benefits including high binding capacity at high flow rates, and scalability.

Each Adenopump™ Adenovirus Purification Unit is capable of purifying up to 5×10^{12} viral particles in less than two hours.

ADENOPURE™ KIT COMPONENTS

There are six (6) components included in this Adenopure™ Kit. Prior to use, please familiarize yourself with each component and its use. If any component is missing or damaged, contact Puresyn's Technical Support at 888-245-9182 for replacement.

- 10X Dilution Buffer (50ml)
- Equilibration-Wash Buffer (270ml)
- Elution Buffer (30ml)
- Benzonase® Stock (4000 U)
- 10cc Syringes (3)
- Adenopump™ Adenovirus Purification Units (3)

ADDITIONAL MATERIALS REQUIRED

- Low protein binding 0.2µm filter units with glass fiber pre-filters (Example: Millipore 0.22µm Durapore PVDF membrane Stericup™ filter unit with a glass fiber pre-filter; catalog numbers SCGV U01 RE and AP20 075 00).
- 15ml and 50ml sterile conical tubes
- Ring-stand/clamp
- Centrifuge
- 37°C water bath or incubator
- Sterile 100-200ml bottles
- Ethanol/dry ice bath

SHIPPING AND STORAGE

The Adenopure™ Kit is shipped at ambient temperature. Upon arrival, store the supplied stock of Benzonase® at -20°C. Store all other Adenopure™ Kit components at room temperature. The shelf life of the Adenopure™ Kit when stored properly is at least 12 months.

ADENOVIRUS PROPAGATION GUIDELINES

The following discussion of adenovirus propagation is for guideline purposes only. Specific adenovirus constructs may need to be propagated differently.

Wild type adenovirus or adenovirus recombinants are usually propagated in the HEK293 cell line (Graham, et al.). HEK293 cells are maintained in a 37°C, 5% CO₂ incubator in DME medium supplemented with 10% fetal bovine serum.

For viral propagation, HEK293 cells may be seeded into 150cm² plates or flasks. When the HEK293 cells reach a confluency of approximately 80% they are infected with the adenovirus stock of interest at multiplicity of infection of 100 (as determined by HPLC analysis or OD_{260nm} readings) in DME medium supplemented with 2% fetal bovine serum (20ml/plate or flask). Infected cells are maintained in a 37°C, 5% CO₂ incubator until harvest. For an average yielding vector, five (5) 150cm² plates or flasks of infected cells will generally yield an adequate amount of adenovirus for one (1) purification procedure utilizing the Adenopure™ Kit (approximately 1×10^{12} to 5×10^{12} viral particles per 100ml infected cell suspension).

Infected cells are harvested 2 to 5 days post-infection (usually when full cytopathic effect is observed). Virus is released by subjecting the infected HEK293 cell suspensions to three (3) freeze-thaw cycles alternately using a 37°C water bath and an ethanol/dry ice bath. Additional freeze-thaw cycles are not recommended as this may lead to adenovirus degradation.

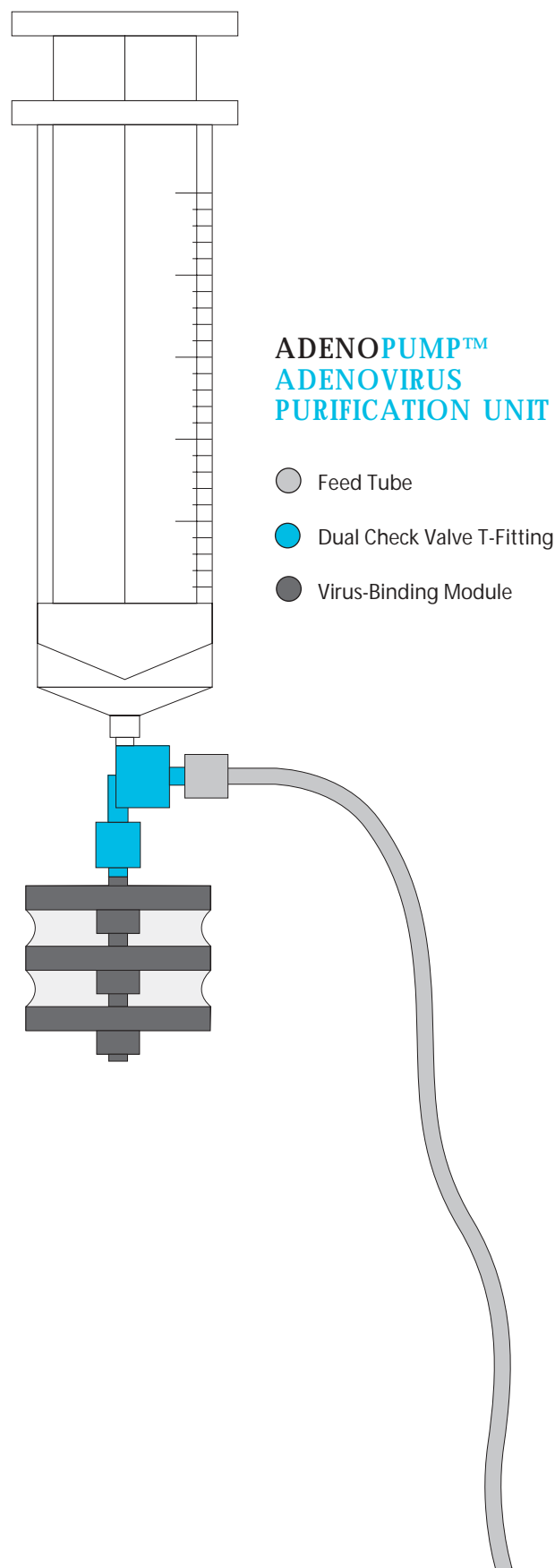
CONTRAINDICATIONS OF ADENOPURE™ KIT USE

As with any chromatographic purification process, Adenopure™ kit purification yield will suffer in severe under-load conditions. For the Adenopure™ kit to function efficiently, a minimum of 1×10^{12} viral particles should be present per 100ml of infected cell suspension.

The Adenopure™ kit has been optimized for adenovirus type 5 vector purification, and the performance of the kit with non-adenovirus type 5 constructs cannot be predicted or guaranteed. The performance of the Adenopure™ kit with adenovirus type 5 vectors that have modifications in viral capsid proteins resulting in a change in the charge density of the virions also cannot be predicted or guaranteed.

ADENOPUMP™ ADENOVIRUS PURIFICATION UNIT OPERATION

1. Mount the Adenopump™ in a clamp attached to a ring-stand.
2. Make sure all the connections of the Adenopump™ are tight.
3. Fully depress the syringe plunger.
4. Place the *feed tube* into a reservoir containing the solution to be passed over the *virus-binding module*.
Note: Do not place the *feed tube* directly into any of the supplied buffer containers because cross contamination may occur between adenovirus purification procedures. When purification steps require it, pour the appropriate amount of supplied buffer into a clean feed reservoir.
5. Pull up on the syringe plunger drawing the solution through the *one-way dual check valve T-fitting* into the syringe. Push down on the syringe plunger to pass the solution back through the *one-way dual check valve T-fitting* and over the *virus-binding module*. Repeat the process as many times as necessary to pass all of the solution over the *virus-binding module*. Try not to push air through the *virus-binding module* as this can prevent even distribution of solution across the membranes in the module.
6. For the virus elution step, detach the *virus-binding module* from the *one-way dual check valve T-fitting* of the Adenopump™, and reattach the module to a 10cc syringe containing Elution Buffer.
7. Optimum Flow Rate: In general, slower is better with a flow rate of 10ml/minute being suitable. A flow rate of 10ml/minute may be approximated if you can "count the drops" as they come out of the *virus-binding module*. A flow rate that results in a stream of material coming out of the *virus-binding module* is too fast. If you are unsure how fast you are pushing material through the purification unit, a direct measure may be made with a timer, and a disposable 15ml conical tube.



2-5 DAY INFECTED CELL SUSPENSION SAMPLE PREPARATION

1. Collect the infected cells and media from five (5) 150cm² tissue culture plates or flasks. If necessary, detach infected cells from the tissue culture plates or flasks by pipetting or by using a cell scraper.

Note: For an average yielding vector, five (5) 150cm² plates or flasks of infected cells will generally yield an adequate amount of adenovirus for one (1) purification procedure utilizing the Adenopure™ Kit (approximately 1×10^{12} to 5×10^{12} viral particles per 100ml infected cell suspension).

2. Process the cell suspension through three (3) freeze-thaw cycles alternately using a 37°C water bath and an ethanol/dry ice bath.
Note: Additional freeze-thaw cycles are not recommended as this may lead to adenovirus degradation.

3. Pellet cell debris by centrifugation at 2000xg for 5 minutes at room temperature.

4. Filter the adenovirus containing supernatant through a 0.2µm filter unit with glass fiber pre-filter. Pre-wetting the filter is usually unnecessary.

5. Add 50µl of 25U/µl Benzonase® to the filtered lysate and mix by gently swirling. Incubate the mixture at 37°C for 30 minutes.

Note: The amount of Benzonase® added is dependant on the overall amount of substrate, not substrate concentration.

6. **Dilute:** Add 10X Dilution Buffer to the Benzonase®-treated filtered lysate and mix by gentle swirling. Determine the amount of 10X Dilution Buffer required using the following formula:

Volume of filtered cell lysate ÷ 9 = Volume of 10X Dilution Buffer required.

Note: Accurate sample volume determination is critical, as incorrect volume addition of 10X Dilution Buffer will lead to poor binding of adenovirus to the *virus-binding module* of the syringe-check valve purification unit.

7. Proceed to adenovirus purification.

ADENOVIRUS PURIFICATION

1. Fill one of the supplied 10cc syringes with 3ml of Elution Buffer and 7ml of air. Set aside for the elution step.
2. **Equilibrate:** Equilibrate the *virus-binding module* of an Adenopump™ by passing 30ml of Equilibration-Wash Buffer through the unit.
Notes: a) Avoid pushing air through the *virus-binding module* as this can prevent even distribution of solution across the membranes in the module. Leaving a small amount of Equilibration-Wash Buffer in the syringe when performing step 3 will not interfere with efficient adenovirus purification; b) Do not place the *feed tube* directly into any of the supplied buffer containers because cross contamination may occur between adenovirus purification procedures. When purification steps require it, pour the appropriate amount of supplied buffer into a clean feed reservoir.
3. **Load:** Load the diluted-filtered lysate onto the *virus-binding module* using the Adenopump™. Repeat the loading step as many times as required to pass the entire sample over the *virus-binding module*.
Note: In general, a slower flow rate will increase the efficiency of binding of adenovirus to the *virus-binding module* with a flow rate of 10ml/minute being suitable.
4. **Wash:** Wash the adenovirus bound to the *virus-binding module* by passing 50ml of Equilibration-Wash buffer over the module using the Adenopump™.
Notes: a) Avoid pushing air through the *virus-binding module* as this can prevent even distribution of solution across the membranes in the module; b) Do not place the *feed tube* directly into any of the supplied buffer containers because cross contamination may occur between adenovirus purification procedures. When purification steps require it, pour the appropriate amount of supplied buffer into a clean feed reservoir.
5. **Elute:** Detach the *virus-binding module* from the *one-way dual check valve T-fitting* of the Adenopump™, and reattach the module to the 10cc syringe containing Elution Buffer prepared in step 1. Collect all material at this step as one fraction as it contains the purified

adenovirus. Pass 15 drops of Elution Buffer through the *virus-binding module*. Incubate the *virus-binding module* for 5 minutes at room temperature before continuing. Pass the remainder of the Elution Buffer through the *virus-binding module* and push the air in the syringe through as well to expel all remaining Elution Buffer from the module.

Notes: a) During the 5 minute incubation, the syringe virus-binding module may be laid on its side or held vertically; b) The final elution volume will be approximately 4.5ml if 3ml of Elution Buffer was utilized in the elution step. The volume increase is due to approximately 1.5ml of residual Wash Buffer remaining in the virus-binding module from the Wash step.

POST-PURIFICATION GUIDELINES

Adenopure™ purified adenovirus preparations should be 0.2µm syringe-filtered prior to use in any tissue culture or animal experiments. Adenopure™ purified adenovirus preparations may be stored at 4°C for short-term use (up to one week), or at -20°C for long-term storage. Repeated freeze-thaw cycles are not recommended.

If a particular storage or formulation buffer is desired for the final adenovirus preparation, the purified adenovirus may be dialyzed into the buffer of interest. A small-scale tangential flow filtration (TFF) device may also be used for buffer exchange and/or additional sample concentration.

QUANTITATION OF ADENOVIRUS

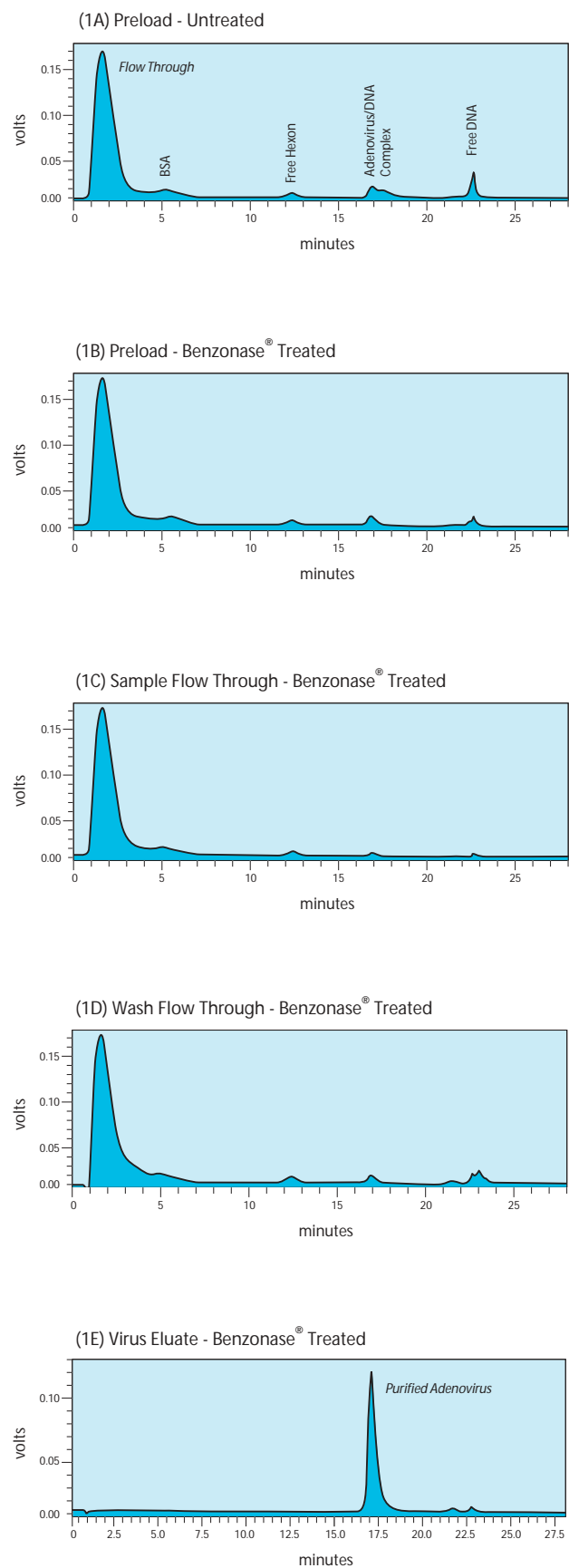
Purified adenovirus may be quantitated using several methods including HPLC (Shabram et al.) and UV spectrophotometry (Maizel et al.). Biological activity may be quantitated using several methods including plaque and CPE assays and gene-expression specific bioassays.

SCIENTIFIC SUPPORT

Successful purification of adenovirus from crude cell lysate is challenging. Crude cell lysate typically contains contaminating cellular proteins and nucleic acids as well as excess viral components not incorporated into intact progeny virions. Additionally, purification processes must deal with typical tissue culture media additives such as fetal bovine serum components.

A series of HPLC chromatograms of fractions collected during the purification process are presented in **Figure 1** to illustrate the functionality of the Adenopure™ Kit. **Figure 1A** shows an HPLC chromatogram of adenovirus-infected crude cell lysate. Under the chromatographic conditions employed in this analysis, peaks resolve for cellular proteins and other debris (flow through), BSA, free adenovirus hexon, adenovirus, adenovirus-DNA complex and free DNA. **Figure 1B** shows that upon Benzonase® treatment of the crude cell lysate, the adenovirus-DNA complex resolves into a peak enriched for free adenovirus. The free DNA peak is also significantly reduced. **Figure 1C** shows that the majority of free cellular proteins, BSA, free adenovirus hexon and free DNA do not bind to the *virus-binding module* of the Adenopump™ under the buffer conditions utilized, and are present in the sample flow through fraction. **Figure 1D** shows that the remainder of free cellular proteins, BSA, free adenovirus hexon and free DNA are removed from the *virus-binding module* of the Adenopump™ during the wash step under the buffer conditions utilized. **Figure 1E** shows the eluted adenovirus as a uniform peak free from other contaminants.

Figure 1: Adenopure™ Kit HPLC Chromatograms



TROUBLESHOOTING

Problem	Cause	Solution
No virus yield	Adenopure™ procedure was not followed	Read and follow Adenopure™ instructions
	Virus stock used for infection is not viable	Obtain a viable virus stock
Low virus yield	Adenopump™ under-loaded due to low virus yield	Re-optimize virus production
	Flow rate through Adenopump™ too fast	Slow flow rate down to 10ml/minute
	Air pushed through virus-binding module during purification	Avoid pushing air through the virus-binding module
	Elution step performed too fast	Slow flow rate down during elution step
	Wrong buffers used during procedure	Read and follow Adenopure™ instructions
	Benzonase® under- or over-digestion	Read and follow Adenopure™ instructions
	Buffer left in virus-binding module during elution	Collect all buffer from virus-binding module during elution
Virus-binding module clogs	Too much residual cellular debris	Pass cell lysate through a 0.2µm filter prior to purification
	Too much residual cellular DNA	Follow instructions for Benzonase® digestion step
Purified adenovirus contains a high level of cellular DNA	Benzonase® treatment incomplete, due to incorrect incubation time or temp.	Follow instructions for Benzonase® digestion step
	Extract from too many plates of infected cells being purified	Lower the amount of extract being purified to (5) 150cm ² plate equivalents
	Benzonase® stock inactive	Store Benzonase® at recommended temperature
Leaks in Adenopump™	Junctions loose in Adenopump™	Tighten all junctions of the Adenopump™ prior to use

CURRENT PURIFICATION PRODUCTS FROM PURESYN

- Adenopure™ Adenovirus Purification Kit
- Adenopure LS™ Large Scale Adenovirus Purification Kit
- PolyFlo® Resin
- Pre-packed columns with PolyFlo® resin designed for small to gram-scale plasmid purification
- Contract services for fermentation, supercoiled plasmid purification, RNA purification and oligonucleotide purification
- Adenoserve™ contract services for adenovirus purification

FUTURE PURIFICATION PRODUCTS FROM PURESYN

- RNA
- AAV Kit

Visit our website for product updates at www.puresyn.com

Technical Support

Representatives from Puresyn are available via phone or e-mail for technical support Monday through Friday from 8:30 am to 4:30 pm Eastern Standard Time. Please call 888-245-9182 and ask for technical support, or write to:

Technical Support
Puresyn, Inc.
87 Great Valley Parkway
Malvern, PA 19355

You may also contact us via fax at 610-640-0808, or e-mail Adenopure@puresyn.com

Important Notice

The components of this adenovirus purification kit are intended for **research laboratory use only** and should not be used for any human or animal diagnostic or therapeutic application.

Warning: Utilization of this Kit exposes the user to potentially hazardous biological agents, i.e. recombinant adenovirus, and may generate biohazardous waste material. Wear safety glasses, gloves, and a lab coat when using this Kit. Follow established safety protocols for handling of biological agents and disposal of biohazardous waste as determined by federal, state and local regulations. It is the user's responsibility to consult the applicable MSDS(s) before using this Kit.

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Graham, F. L., Smiley, J., Russel, W. C., and Narin, R. (1977). Characteristics of a human cell line transformed by DNA from human adenovirus type 5. *J. Gen. Virol.* **36**, 59-72

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Trademarks and Patents

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Acknowledgements

The cover figure shows the three-dimensional structure of the human adenovirus type 5 virion at 15Å resolution, as obtained by three-dimensional reconstruction from cryo-electron microscopy images. Only short portions of the fibers are properly reconstructed because the fibers do not follow icosahedral symmetry. For a review of the methodologies utilized in three-dimensional reconstruction of icosahedral viruses, see Baker et al. Figure courtesy of Carmen San Martin, Ph.D., and Roger M. Burnett, Ph.D., The Wistar Institute, 3601 Spruce Street, Philadelphia, PA 19104.

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